

PureLyse™ Protocol for Cultured Cells (For research use only)

1. Set-up

- a. Harvest cells in a microcentrifuge tube by centrifuging.
- b. Discard supernatant and resuspend pelleted cells with 500 µl 1X *Binding Buffer*.
- c. Attach an empty 1 ml syringe to the Luer fitting of the PureLyse™ cartridge. Attach the battery pack to the connector of the PureLyse™ cartridge.

2. Lysis and Extraction

- a. Withdraw sufficient sample volume through the tip of the device to fill the chamber (~ 125 µl).
- b. Turn on the device. Draw the remaining sample through the PureLyse™ cartridge into the syringe. Be sure that the cartridge chamber remains filled with sample.
- c. Reverse the direction of the syringe, dispensing the sample back through the cartridge into the original sample container. Again, the cartridge chamber should remain filled with sample.
- d. Continue withdrawal and infusion of sample for 2.5 minutes total (~ 10 to 15 passes).
- e. After the final pass, dispense the entire lysate into the sample tube, emptying the chamber). Use additional air to completely purge the sample from the cartridge. Turn off the device.

3. Elution

- a. Transfer 125 µl 1x *Elution Buffer* to a new microfuge tube.
- b. Attach a clean 1 ml syringe to the PureLyse™ cartridge.
- c. Place the tip of the device into the tube containing the elution buffer and withdraw the 125 µl of 1x *Elution Buffer* to fill the chamber.
- d. Turn on the device and allow to mix for 1 minute.
- e. Dispense the eluate back into the tube. Use additional air to completely purge the sample from the cartridge. Turn off the device. For additional yield, another elution step may be performed using an additional 125 µl 1x *Elution Buffer*.

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